

P. Pankajakshan, *et al.*

Point-spread function model for fluorescence MACROscopy imaging

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Road map

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Figure 1: Leica MACROFLuoTM



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Figure 1: Leica MACROFLuoTM

What is claimed?

▶ large object fields (up to 35mm),



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Figure 1: Leica MACROFLuoTM

What is claimed?

- ▶ large object fields (up to 35mm),
- ▶ large working distances (up to 97mm),



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Figure 1: Leica MACROFLuoTM

What is claimed?

- ▶ large object fields (up to 35mm),
- ▶ large working distances (up to 97mm),
- parallax-free and precise imaging,



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Figure 1: Leica MACROFLuoTM

What is claimed?

- ▶ large object fields (up to 35mm),
- ▶ large working distances (up to 97mm),
- parallax-free and precise imaging,
- multi-color fluorescence at high resolution.





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Biological specimens often scatter light,





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- Biological specimens often scatter light,
- image living samples without the need for physical sectioning,





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- Biological specimens often scatter light,
- image living samples without the need for physical sectioning,
- image deeper into the sample,





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- Biological specimens often scatter light,
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- image deeper into the sample,
 - spherical aberrations due to refractive index mismatch deteriorate image quality,





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- Biological specimens often scatter light,
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 - physical limitation of depth of field.





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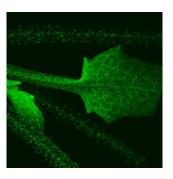


Figure 2: Confocal MACROscope image of *Arabidopsis Thaliana* seedlings ©INRA Sophia Antipolis, France.



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 $NA = n_i \sin \theta_{max}$



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$$NA = n_i \sin \theta_{\text{max}}$$
$$= \sin \theta_{\text{max}}$$



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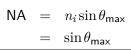
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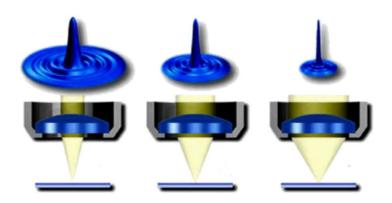


Figure 3: Numerical aperture and resolution for microscope.



Numerical aperture and working distance

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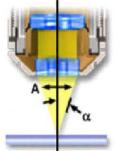




Figure 4: Schema of numerical aperture influencing working distance.



Why do we need a point-spread function (PSF)?

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Figure 5: Confocal MACROscope image of *Arabidopsis Thaliana* seedlings ©INRA Sophia Antipolis, France.



Experiments on the MACROscope

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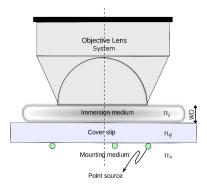


Figure 6: Experimental determination of the impulse response of the MACROscope by imaging point sources.



Experiments on the MACROscope

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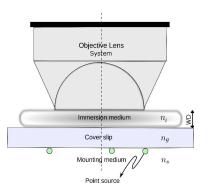


Figure 6: Experimental determination of the impulse response of the MACROscope by imaging point sources.

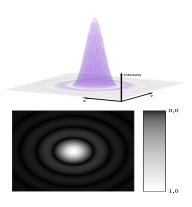


Figure 7: 2D Airy disk function.



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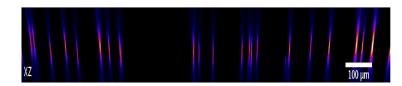


Figure 8: Radial projection of $4\mu m$ beads as imaged from a Leica Widefield MacroFluoTMZ16 APO. The objective has a 5X magnification and a zoom of up to 16X. ©Herbomel lab, Pasteur Institute.



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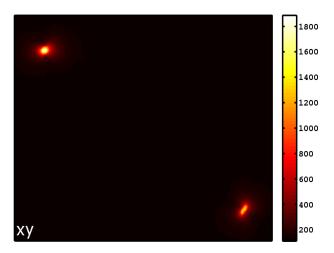


Figure 9: Axial projection of $4\mu m$ beads as imaged from a Leica Widefield MacroFluoTMZ16 APO. The objective has a 5X magnification and a zoom of 1.6X. ©Herbomel lab, Pasteur Institute.



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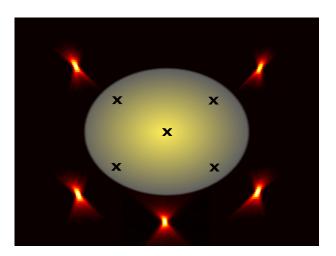


Figure 10: Bead images taken at different lateral positions of the field (as marked by the cross).



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When the acquisition parameters are known, restoration can be achieved by using either theoretically calculated or estimated PSFs.



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When the acquisition parameters are known, restoration can be achieved by using either theoretically calculated or estimated PSFs.

▶ If $P(k_x, k_y, z)$ is the 2D complex pupil function and λ is the wavelength, the amplitude PSF can be calculated by just $2N_z$ number of 2D FFTs

$$h_A(x, y, z; \lambda) = \int_{k_x} \int_{k_y} P(k_x, k_y, z) \exp(j(k_x x + k_y y)) dk_y dk_x$$



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- When the acquisition parameters are known, restoration can be achieved by using either theoretically calculated or estimated PSFs.
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▶ and the incoherent PSF is

$$h_{Th}(\mathbf{x}; \lambda_{\mathsf{ex}}, \lambda_{\mathsf{em}}) = C|h_A(\mathbf{x}; \lambda_{\mathsf{ex}})| \times |h_A(\mathbf{x}; \lambda_{\mathsf{em}})|$$



Asilomar coffee cup

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Figure 11: Out-of-focus highlights (OOFH) for the Asilomar coffee cup. (Photograph by Praveen Pankajakshan.)



Cat'e eye effect

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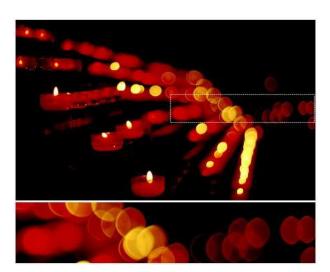


Figure 12: The Cat's eye effect is seen in the out-of-focus highlights (OOFH). (Photograph by Peter Boehmer.)

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Optical vignetting

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Figure 13: Lens viewed from the front. (Photograph by Peter Boehmer.)



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Figure 13: Lens viewed from the front. (Photograph by Peter Boehmer.)



Figure 14: Lens viewed from the side. (Photograph by Peter Boehmer.)



Limiting apertures overlapping

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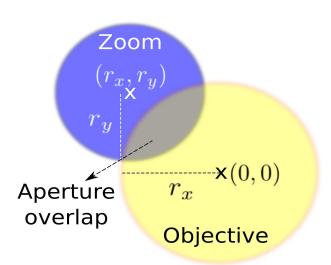
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Pacific grove beach at no zoom

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Figure 15: Optical vignetting example at minimum zoom. (Photograph by Praveen Pankajakshan.)



Pacific grove beach zoom position $1\times$

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Figure 16: Optical vignetting disappears as zoom is increased. (Photograph by Praveen Pankajakshan.)



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► For a MICROscope, the pupil function is



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► For a MICROscope, the pupil function is

$$P_m(k_x,k_y,z;\lambda) = \begin{cases} e^{jk_0\phi(\theta_i,\theta_s,z)}, & \text{ if } \sqrt{k_x^2 + k_y^2} < \frac{2\pi}{\lambda} \mathsf{NA}_{\mathsf{Obj}} \\ 0, & \text{ otherwise.} \end{cases}$$



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► For a MICROscope, the pupil function is

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modification for a MACROscope

$$P(k_x,k_y,z;\lambda) = \begin{cases} P_{m}, & \text{ if } \sqrt{(k_x-r_x)^2+(k_y-r_y)^2} < \frac{2\pi}{\lambda} \mathsf{NA}_{\mathsf{Zo}} \\ 0, & \text{ otherwise}. \end{cases}$$



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► For a MICROscope, the pupil function is

$$P_m(k_x,k_y,z;\lambda) = \begin{cases} e^{jk_0\phi(\theta_i,\theta_s,z)}, & \text{ if } \sqrt{k_x^2 + k_y^2} < \frac{2\pi}{\lambda} \mathsf{NA}_{\mathsf{Obj}} \\ 0, & \text{ otherwise.} \end{cases}$$

modification for a MACROscope

$$P(k_x,k_y,z;\lambda) = \begin{cases} \textcolor{red}{P_m}, & \text{ if } \sqrt{(k_x-r_x)^2+(k_y-r_y)^2} < \frac{2\pi}{\lambda} \mathsf{NA}_{\mathsf{Zo}} \\ \textcolor{red}{0}, & \text{ otherwise}. \end{cases}$$

parameters of the PSF: $\omega_h = \{NA_{Obj}, NA_{Zo}, r_x, r_y\}$



Compare experimental PSF and theoretical PSF

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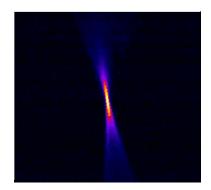


Figure 17: Experimental image of point source. Bead diameter 4μ m and zoom 1X. ©Imaging Center, IGBMC, France.



Compare experimental PSF and theoretical PSF

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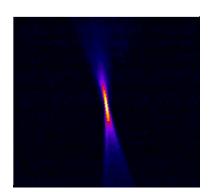


Figure 17: Experimental image of point source. Bead diameter 4μ m and zoom 1X. ©Imaging Center, IGBMC, France.

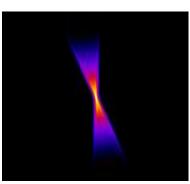


Figure 18: Simulated image of point source assuming the maximum NA of 0.5.



Optically vignetting the pupil

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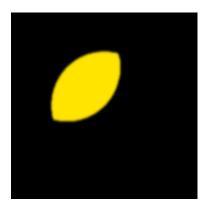


Figure 19: Simulating the vignetting.



Optically vignetting the pupil

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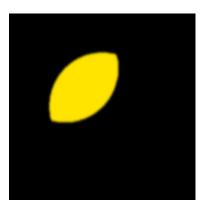
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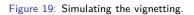
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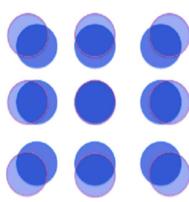


Figure 20: Schematic of overlapping apertures.



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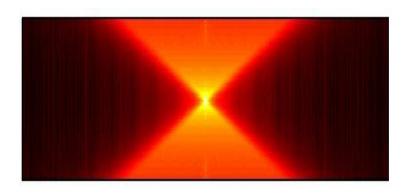


Figure 21: Simulated image of point source assuming 100% overlap of the apertures.



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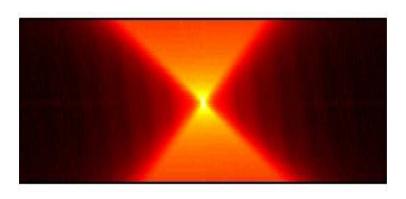


Figure 22: Simulated image of point source assuming an overlap of 75% of the apertures.



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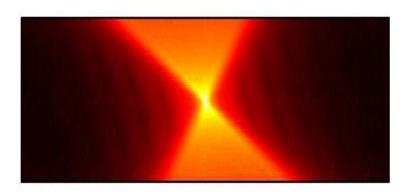


Figure 23: Simulated image of point source assuming an overlap of 50% of the apertures.



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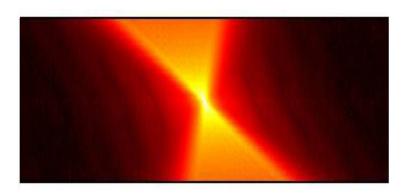


Figure 24: Simulated image of point source assuming an overlap of 25% of the apertures.



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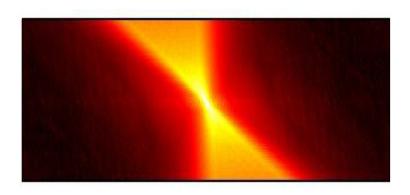


Figure 25: Simulated image of point source assuming an overlap of 10% of the apertures.



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► MACROscope PSF varies as a function of the lateral position,



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- MACROscope PSF varies as a function of the lateral position,
- vignetting was observable only for smaller zooms (large FOV),



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- MACROscope PSF varies as a function of the lateral position,
- vignetting was observable only for smaller zooms (large FOV),
- the initial PSF model will be enhanced further with newer acquisitions on different systems,



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- ► MACROscope PSF varies as a function of the lateral position,
- vignetting was observable only for smaller zooms (large FOV),
- the initial PSF model will be enhanced further with newer acquisitions on different systems,
- ► restoration of images can be achieved with the complete or partial knowledge of the theoretical PSF.



Acknowledgements

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- ► The first author wish to thank ANR DIAMOND for funding the postdoctoral research fellowship,
- ▶ the authors are grateful to Dr. Philippe Herbomel from the Institut Pasteur, France and Dr. Didier Hentsch from IGBMC, France for the images and the discussions,
- ▶ some of the slides in this presentation were prepared by Dr. Mickael Lelek from Institut Pasteur, France, a big thanks to him,
- we acknowledge our colleagues for their support.

For more information see: http://www-syscom.univ-mlv.fr/ANRDIAMOND/.